Research Article

Synthesis of ¹³C and ¹⁵N multilabeled 5-aminolevulinic acid

Katsumi Iida and Masahiro Kajiwara* Department of Medicinal Chemistry, Meiji Pharmaceutical University, 2-522-1 Noshio, Kiyose-shi, Tokyo 204-8588, Japan

Summary

 $5-[4-{}^{13}C, {}^{15}N]-$ and $5-[5-{}^{13}C, {}^{15}N]$ Aminolevulinic acid (ALA) were simply synthesized in four steps by the condensation of $[1-{}^{13}C, {}^{15}N]-$ or $[2-{}^{13}C, {}^{15}N]$ glycine, respectively, with phthalic anhydride, followed by conversion to the chloride, coupling reaction with a three-carbon unit and hydrolysis. Copyright @ 2002 John Wiley & Sons, Ltd.

Key Words: $5-[4-{}^{13}C, {}^{15}N]$ aminolevulinic acid; $5-[5-{}^{13}C, {}^{15}N]$ aminolevulinic acid; $[1-{}^{13}C, {}^{15}N]$ glycine; $[2-{}^{13}C, {}^{15}N]$ glycine

Introduction

We have developed various methods for the synthesis of regioselectively stable-isotope-labeled 5-aminolevulinic acid (ALA),^{1,2} which is a precursor of porphyrins, for use in studies on the biosynthesis of porphyrins.^{3–5} Synthetic methods of ALA isotope-labeled on C-5 have been reported by Shemin *et al.*,⁶ Battersby *et al.*⁷ and from our laboratory,¹ but involve many steps between introduction of the labeled carbon and formation of the final product. We present here a simple synthesis of $[5-{}^{13}C, {}^{15}N]ALA$ from $[2-{}^{13}C, {}^{15}N]$ glycine, and of $[4-{}^{13}C, {}^{15}N]$ ALA from $[1-{}^{13}C, {}^{15}N]$ glycine.

*Correspondence to: M. Kajiwara, Department of Medicinal Chemistry, Meiji Pharmaceutical University, 2-522-1 Noshio, Kiyose-shi, Tokyo 204-8588, Japan.

Copyright © 2002 John Wiley & Sons, Ltd.

Received 9 October 2001 Accepted 24 October 2001

Results and discussion

As shown in Scheme 1, the condensation of $[2^{-13}C, {}^{15}N]$ glycine (1) and phthalic anhydride (2) gave 2- $[{}^{15}N]$ phthalimido $[2^{-13}C]$ acetic acid (3) in 86% yield. The transhalogenation of ethyl 3-bromopropionate (5) with sodium iodide in 2-butanone gave ethyl 3-iodopropionate (6) in 95% yield. The coupling reaction^{8,9} of 2- $[{}^{15}N]$ phthalimido $[2^{-13}C]$ acetyl chloride (4), which was derived from 2- $[{}^{15}N]$ phthalimido $[2^{-13}C]$ acetic acid (3) and thionyl chloride, with 2-ethoxycarbonylethylzinc iodide, which was derived from ethyl 3-iodopropionate (6) treated with zinccopper couple, catalyzed by tetrakis(triphenylphosphine)palladium (0) in dry toluene and dry *N*,*N*-dimethylacetamide gave ethyl 5- $[{}^{15}N]$ phthalimido $[5^{-13}C]$ levulinate (7) in 92% yield. Finally, hydrolysis of ethyl 5- $[{}^{15}N]$ phthalimido $[5^{-13}C]$ levulinate (7) in 6 N hydrochloric acid gave $[5^{-13}C, {}^{15}N]$ ALA (8) in 89% yield. $[4^{-13}C, {}^{15}N]$ ALA was similarly synthesized from $[1^{-13}C, {}^{15}N]$ glycine in four steps.

Experimental

Materials

 $[1-{}^{13}C, {}^{15}N]$ - and $[2-{}^{13}C, {}^{15}N]$ Glycine (99 at% ${}^{13}C, 99$ at % ${}^{15}N$) were purchased from Masstrace, Inc. All other chemicals were of analytical grade.

Instruments

Melting point determinations were carried on a Yanaco micro melting point apparatus, Model MP; values are uncorrected. IR spectra were



Scheme 1. Synthesis of [5-¹³C,¹⁵N]ALA (8)

Copyright © 2002 John Wiley & Sons, Ltd.

J Label Compd Radiopharm 2002; 45: 139-143

recorded on a Jasco VALOR-III FT-IR spectrometer. ¹H-NMR (300 MHz) and ¹³C-NMR (75 MHz) spectra were recorded on a Varian GEMINI-300 spectrometer. EI- and FAB-MS spectra were obtained on a Jeol DX302 spectrometer.

2-[¹⁵N]Phthalimido[2-¹³C]acetic acid (3)

[2-¹³C, ¹⁵N]Glycine (1) (1.0 g, 13.0 mmol) and phthalic anhydride (2) (2.0 g, 13.5 mmol) were heated at 160°C, until the whole was melted. After cooling, the residue was recrystallized from water to give 2-[¹⁵N]phthalimido[2-¹³C]acetic acid (3) (2.3 g, 86%), m.p. 193–195°C; ¹H-NMR (CDCl₃) δ : 4.49 (dd, 2H, J = 142.3 Hz, ²J = 1.1 Hz), 7.75–7.92 (m, 4H); ¹³C-NMR (CDCl₃) δ :38.4 (d, J = 13.3 Hz); FT-IR (KBr) cm⁻¹: 1745, 2928; EI-MS m/z (rel. int. %): 207 (M⁺, 4), 162 (100).

Analytical data of 2-[¹⁵N]phthalimido[1-¹³C]acetic acid, m.p. 191–194°C; ¹H-NMR (CDCl₃) δ :4.50 (dd, 2H, ²J=5.8 Hz, ²J=1.1 Hz), 7.75–7.92 (m, 4H); ¹³C-NMR (CDCl₃) δ :171.6; FT-IR (KBr) cm⁻¹; 1726, 2926; EI-MS m/z (rel. int. %): 207 (M⁺, 4) 161 (100).

Ethyl 3-iodopropionate (6)

A suspension of ethyl 3-bromopropionate (5) (10 ml, 78.0 mmol) and NaI (17 g, 113.4 mmol) in 2-butanone (100 ml) was refluxed at 90°C for 16 h under argon. After cooling, the crystals were removed by filtration. The filtrate was evaporated, diluted with ether, and washed with brine. Distillation of the crude product gave ethyl 3-iodopropionate (6) (33.7 g, 95%), b.p. 105°C (20 mm Hg); ¹H-NMR (CDCl₃) δ :1.29 (t, 3H, J = 7.1 Hz), 2.97 (t, 2H, J = 7.1 Hz), 3.34 (t, 2H, J = 7.1 Hz), 4.19 (q, 2H, J = 7.1 Hz); FT-IR (neat) cm⁻¹: 1730; EI-MS m/z (rel. int. %): 228 (M⁺, 100), 155 (61), 101 (92), 73 (47).

Ethyl 5-[¹⁵N]phthalimido[5-¹³C]levulinate (7)

A mixture of 2-[¹⁵N]phthalimido[2-¹³C]acetic acid ($\underline{3}$) (2.3 g, 11.1 mmol) and thionyl chloride (30 ml) was refluxed at 80°C for 18 h. The excess thionyl chloride was removed *in vacuo*. The residue was dissolved in dry

Copyright © 2002 John Wiley & Sons, Ltd. J Label Compd Radiopharm 2002; 45: 139-143

toluene (3 ml) and evaporated. This process was repeated twice, and the resulting 2-[¹⁵N]phthalimido[2-¹³C]acetyl chloride (4) was dissolved in dry toluene (5 ml) under argon. Ethyl 3-iodopropionate (6) (2.82 g. 12.4 mmol) was added to a suspension of Zn–Cu couple (1.0 g) in dry toluene (1.2 ml) and dry N, N-dimethylacetamide (14 ml) under argon, and the mixture was heated at 60°C for 15h. To this suspension, a suspension of Pd(PPh₃)₄ (530 mg, 459 µmol) in dry toluene (5 ml) under argon was added, and the whole was stirred at 60°C for 5 min. To this suspension, a solution of $2 \cdot [^{15}N]$ phthalimido $[2 - ^{13}C]$ acetyl chloride (4) was added, and the whole was stirred at 60°C for 25 min. The reaction was diluted with ethyl acetate (300 ml), and the whole was washed with 1 N HCl, sat. NaHCO3 and brine, dried over dry MgSO4, and evaporated. Chromatography of the crude product on silica gel with ethyl acetate:hexane (1:1) gave ethyl 5-[¹⁵N]phthalimido[5-¹³C]levulinate (7) (2.98 g, 92%), m.p. 75–77°C; ¹H-NMR (CDCl₃) δ : 1.26 (t, 3H, J = 7.1 Hz), 2.66 (t, 2H, J = 6.6 Hz), 2.85 (t, 2H, J = 6.6 Hz), 4.15 (q, 2H, J = 6.6 Hz), 4. J = 7.1 Hz), 4.56 (d, 2H, J = 139.8 Hz), 7.73–7.89 (m, 4H); ¹³C-NMR (CDCl₃) δ : 46.5 (d, J = 12.2 Hz); FT-IR (KBr) cm⁻¹: 1725, 1771, 2920; EI-MS m/z (rel. int. %): 291 (M⁺, 2), 162 (44), 129 (100), 101 (75).

Analytical data of ethyl 5-[¹⁵N]phthalimido[4-¹³C]levulinate, m.p. 73–76°C; ¹H-NMR (CDCl₃) δ : 1.26 (t, 3H, J=7.1 Hz) 2.67 (q, 2H, J=6.6 Hz), 2.86 (q, 2H, J=6.6 Hz), 4.15 (q, 2H, J=7.1 Hz), 4.57 (dd, 2H, ²J=3.8 Hz, ²J=1.1 Hz), 7.73–7.89 (m, 4H); ¹³C-NMR (CDCl₃) δ : 200.8; FT-IR (KBr) cm⁻¹: 1726, 1770, 2916; EI-MS m/z (rel. int. %): 291 (M⁺, 1), 161 (100), 130 (50), 102 (37).

[5-¹³C,¹⁵N]ALA hydrochloride (8)

A solution of ethyl 5-[¹⁵N]phthalimido[5-¹³C]levulinate (7) (1.38 g, 4.7 mmol) in 6 N HCl (6 ml) was refluxed at 110°C for 14 h. The reaction mixture was cooled to room temperature and filtrated. The filtrate was washed with ether and evaporated. The residue was recrystallized from EtOH–ether to give [5-¹³C,¹⁵N]ALA hydrochloride (8) (717 mg, 89%), m.p. 144–147°C; ¹H-NMR (D₂O) δ : 2.72 (t, 2H, J = 6.2 Hz), 2.90 (t, 2H, J = 6.2 Hz), 4.13 (d, 2H, J = 143.9 Hz); ¹³C-NMR (D₂O) δ : 49.8 (d, J = 7.7 Hz); FT-IR (KBr) cm⁻¹: 1695, 1725, 2927; FAB-MS (glycerol) m/z: 134 (MH⁺-HCl).

Analytical data of $[4^{-13}C, {}^{15}N]ALA$ hydrochloride, m.p. 146–149°C; ¹H-NMR (D₂O) δ : 2.72 (q, 2H, J = 6.0 Hz), 2.90 (q, 2H, J = 6.0 Hz), 4.13 (d, 2H, J = 4.2 Hz); ¹³C-NMR (D₂O) δ : 206.8; FT-IR (KBr) cm⁻¹; 1685, 1725, 3005; FAB-MS (glycerol) m/z: 134 (MH⁺-HCl).

References

- Kurumaya K, Okazaki T, Seido N, Akasaka Y, Kawajiri Y, Kajiwara M, Kondo M. J Labelled Cpd Radiopharm 1989; 27: 217–235.
- Iida K, Takao Y, Ogai T, Kajiwara M. J Labelled Cpd Radiopharm 1997; 39: 797–802.
- Okazaki T, Kurumaya K, Kajiwara M. Chem Pharm Bull 1990; 38: 1727–1730.
- 4. Kajiwara M, Hara K, Mizutani M, Kondo M. *Chem Pharm Bull* 1992; **40**: 3321–3323.
- 5. Spencer JB, Stolowich NJ, Santander PJ, Pichon C, Kajiwara M, Tokiwa S, Takatori K, Scott AI. *J Am Chem Soc* 1994; **116**: 4991–4992.
- 6. Shemin D, Russell CS, Abramsky T. J Biol Chem 1955; 215: 613-626.
- Battersby AR, Hunt E, McDonald E, Moron J. J Chem Soc, Perkin Trans 1 1973: 2917–2922.
- 8. Tamaru Y. Ochiai H, Sanda F, Yoshida Z. Tetrahedron Lett 1985; 26: 5529–5532.
- Tamaru Y, Ochiai H, Nakamura T, Tsubaki K, Yoshida Z. Tetrahedron Lett 1985; 26: 5559–5562.